



**Ascorbic Acid**

**Colorimetric Microplate Assay Kit**

**User Manual**

**Catalog # CAK1048**

(Version 3.1E)

Detection and Quantification of Ascorbic Acid (AsA) Content in  
Tissue extracts, Cell lysate Samples.

**For research use only. Not for diagnostic or therapeutic procedures.**

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## I. INTRODUCTION

Ascorbic Acid, also known as Vitamin C, is a six-carbon lactone produced by plants and some animal species but not by humans and other primates. Ascorbic acid functions as an enzymatic cofactor for multiple enzymes, serving as an electron donor for monooxygenases and dioxygenases. Ascorbic acid also functions as a powerful antioxidant, particularly in regards to reactive oxygen species.

Ascorbic Acid Colorimetric Microplate Assay Kit provides a convenient tool for sensitive detection of ascorbic acid concentration in a variety of samples. The addition of enzyme to parallel samples removes ascorbic acid specifically. Then, the probe is reduced to ferrous ion ( $\text{Fe}^{2+}$ ) by total or residual antioxidant, which reacts with dye reagent to form a colored product. The color intensity reduction after ascorbic acid removal at 510nm is proportional to the ascorbic acid concentration in the sample.

## II.KIT COMPONENTS

| Component         | Volume     | Storage            |
|-------------------|------------|--------------------|
| 96-WellMicroplate | 1 plate    |                    |
| Assay Buffer      | 30 mlx 4   | 4 °C               |
| Reaction Buffer A | 16 ml x 1  | 4 °C               |
| Enzyme            | Powder x 1 | -20 °C             |
| Probe             | Powder x 1 | 4 °C               |
| Reaction Buffer B | 16 ml x 1  | 4 °C               |
| Dye Reagent       | Powder x 2 | 4 °C, keep in dark |
| Standard          | Powder x 2 | 4 °C, keep in dark |
| Technical Manual  | 1 Manual   |                    |

### **III. MATERIALS REQUIRED BUT NOT PROVIDED**

1. Microplate reader to read absorbance at 510 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer
8. Ice

#### IV. REAGENT PREPARATION

**Standard:** Briefly centrifuge prior to opening. Dissolve each vial in 0.25 ml distilled water to generate 50 mmol/L of Stock Solution. Then dilute to 2 mmol/L top standard solution by adding 40  $\mu$ l Stock Solution to 960  $\mu$ l distilled water. Perform 2-fold serial dilutions of the top standard solution using distilled water to make the standard curve. The concentration of standard curve could be 2.0/1.0/0.5/0.25/0.125/0.0625/0.0312 mmol/L. Keep in dark and prepare standard curve fresh for immediate use.

**Enzyme:** Briefly centrifuge prior to opening. Dissolve in 1 ml distilled water before use. Store at -80 °C for 1 month.

**Probe:** Briefly centrifuge prior to opening. Dissolve in 1 ml distilled water to generate probe stock solution. Dilute the stock solution 5-fold using distilled water to prepare the probe working solution (eg. 200  $\mu$ l to 800  $\mu$ l distilled water). Store at -20 °C for 1 month or 4 °C for 3 days.

**Dye Reagent:** Briefly centrifuge prior to opening. Dissolve each vial in 1 ml distilled water before use. Keep in dark and store at -20 °C for 1 month or 4 °C for 3 days.

**Note:** Divide into small aliquots to avoid repeated freeze-thaw cycles.

## V. SAMPLE PREPARATION

### 1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay Buffer for  $5 \times 10^6$  cell or bacteria (the quantity should be adjusted according to the actual situation of the sample), sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 10000g 4°C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

### 2. For tissue samples

Weigh out 0.1 g tissue (the quantity should be adjusted according to the actual situation of the sample), homogenize with 1 ml Assay Buffer on ice, centrifuged at 10000g 4°C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

### 3. For liquid samples

Detect directly or dilute with Assay Buffer.

## VI. ASSAY PROCEDURE

Add samples to a paired set of wells in a 96-well plate as Sample and Control.

Add following reagents into the microplate:

| Reagent*                                                                             | Sample** | Control** | Standard | Blank |
|--------------------------------------------------------------------------------------|----------|-----------|----------|-------|
| Reaction Buffer A                                                                    | 80 µl    | 80 µl     | 80 µl    | 80 µl |
| Sample                                                                               | 10 µl    | 10 µl     | --       | --    |
| Standard                                                                             | --       | --        | 10 µl    | --    |
| Distilled water                                                                      | 10 µl    | --        | 10 µl    | 20 µl |
| Enzyme                                                                               | --       | 10 µl     | --       | --    |
| Mix and put at room temperature for 10 minutes                                       |          |           |          |       |
| Probe                                                                                | 10 µl    | 10 µl     | 10 µl    | 10 µl |
| Reaction Buffer B                                                                    | 80 µl    | 80 µl     | 80 µl    | 80 µl |
| Dye Reagent                                                                          | 10 µl    | 10 µl     | 10 µl    | 10 µl |
| Mix and put in the oven at 37 °C for 5 minutes, record absorbance measured at 510 nm |          |           |          |       |

### Note:

\*Reagents must be added sequentially and should not be premixed prior to addition.

\*\*The concentrations can vary over a wide range depending on the different samples.

For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.

\*\*For colored samples, we recommend setting a parallel sample background control well with same volume of sample only. Other reagents were replaced by distilled water to the same total volume. Subtract the OD value of the sample background control from the OD value of the sample to correct for interference from the sample's own color.



## VII. CALCULATION

1. Calculate the sample concentration in ASSAY PROCEDURE according to the slope of the standard curve

$$C = \frac{(OD_{\text{Sample}} - OD_{\text{Control}}) - \text{Intercept}}{\text{Slope}} \times n (\text{mmol/L})$$

Calculate the initial concentration according to sample preparation procedure.

2. According to one point of the standard OD and concentration

2.1. According to the protein concentration of sample

$$C = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Control}})}{(OD_{\text{Standard}} - OD_{\text{Blank}}) \times (V_{\text{Sample}} \times C_{\text{Protein}})} (\mu\text{mol/mg})$$

2.2. According to the weight of sample

$$C = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Control}})}{(OD_{\text{Standard}} - OD_{\text{Blank}}) \times (W \times V_{\text{Sample}} / V_{\text{Assay}})} (\mu\text{mol/g})$$

2.3. According to the quantity of cells or bacteria

$$C = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Control}})}{(OD_{\text{Standard}} - OD_{\text{Blank}}) \times (N \times V_{\text{Sample}} / V_{\text{Assay}})} (\mu\text{mol}/10^4)$$

Slope: the absorbance slope of standard curve

n: the dilution factor

C<sub>Protein</sub>: the protein concentration of sample, mg/mL

W: the weight of total sample, g

N: the quantity of total cell or bacteria sample, 10<sup>4</sup>

C<sub>Standard</sub>: the concentration of standard, μmol/mL

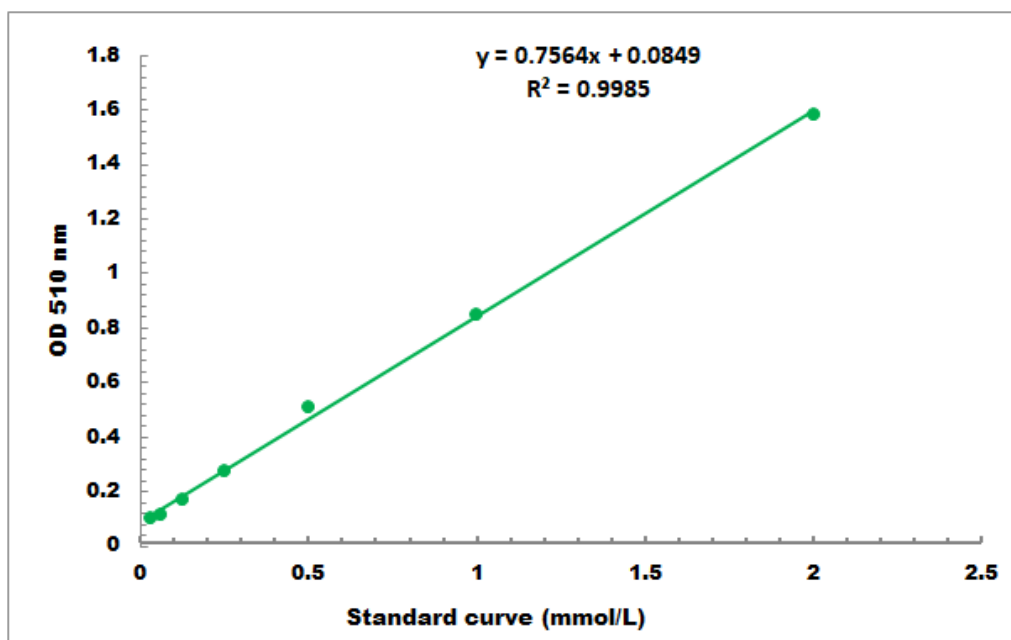
V<sub>Standard</sub>: the volume of standard in assay procedure, mL

V<sub>Sample</sub>: the volume of sample in assay procedure, mL

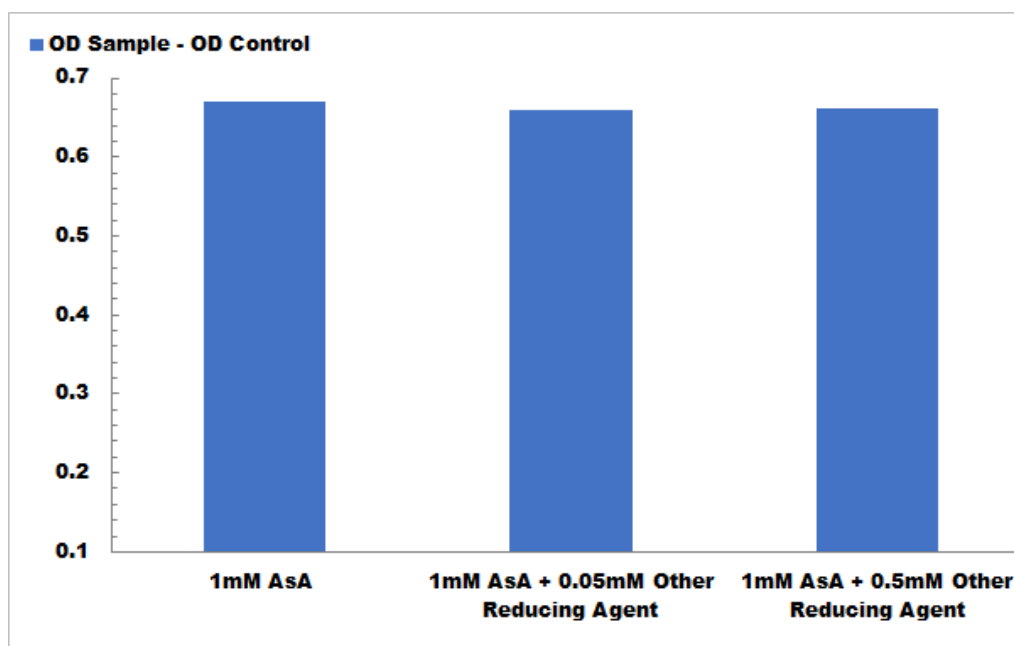
V<sub>Assay</sub>: the volume of Assay Buffer in sample preparation, mL

## VIII. TYPICAL DATA

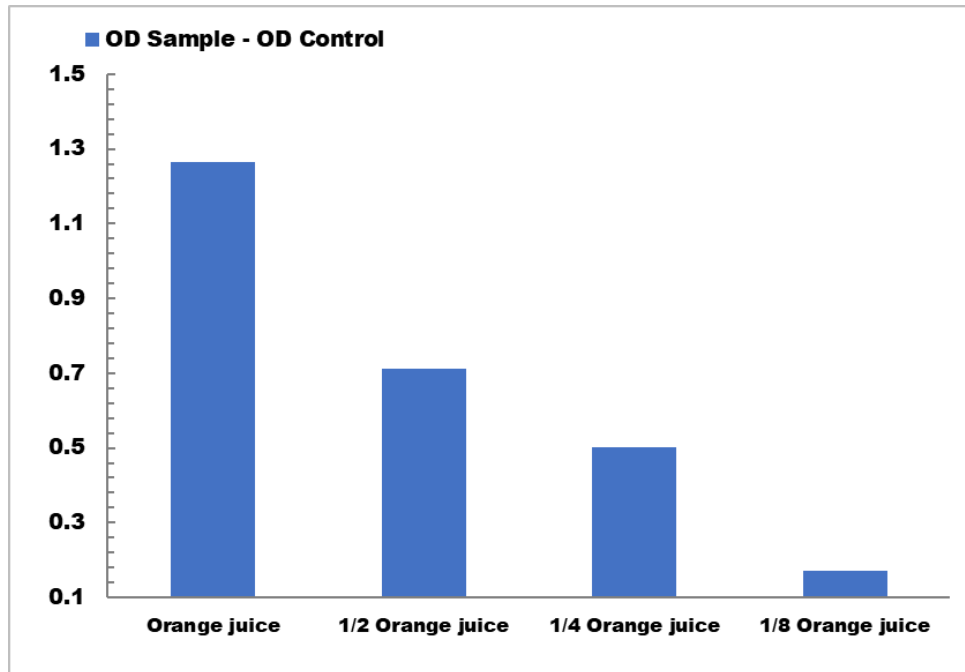
The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.02 mmol/L- 2.0 mmol/L



Detection Ascorbic Acid with other different concentration reducing agent



Determination of ascorbic acid in fruit juices