



L-galactono-1,4-lactone Dehydrogenase Activity Colorimetric Microplate Assay Kit User Manual

Catalog # CAK1050

(Version 1.2C)

Detection and Quantification of L-galactono-1,4-lactone
Dehydrogenase(GalLDH) Activity in Tissue extracts, Cell
lysateSamples.

For research use only. Not for diagnostic or therapeutic procedures.

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I. INTRODUCTION

L-galactono-1,4-lactone Dehydrogenase (EC 1.3.2.3) catalyzes the last step in the main pathway of vitamin C (L-ascorbic acid) biosynthesis in higher plants.

L-galactono-1,4-lactone Dehydrogenase Activity Colorimetric Microplate Assay Kit provides a simple and direct procedure for measuring L-galactono-1,4-lactone Dehydrogenase activity levels. The enzyme catalysed reaction products reduced Cyt c can be measured at a colorimetric readout at 550 nm.

II.KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 mlx 4	4 °C
Substrate I	Powder x 1	4 °C, keep in dark
Substrate II	Powder x 1	4 °C
Technical Manual	1 Manual	

III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 550 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer
8. Ice

IV. REAGENT PREPARATION

Substrate I: Add 17 ml distilled water to dissolve before use. Keep in dark and store at 4 °C for 1-2 weeks or -20°C for 3-4 months after reconstitution.

Substrate II: Briefly centrifuge prior to opening. Add 1 ml distilled water to dissolve before use. Prepare for immediate use or store at -20°C for 3-4 days after reconstitution.

Note: Divide into small aliquots to avoid repeated freeze-thaw cycles.

V. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay Buffer for 5×10^6 cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 13000g 4°C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay Buffer on ice, centrifuged at 13000g 4°C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

VI. ASSAY PROCEDURE

Warm the Substrate I and Substrate II to room temperature before use.

Add following reagents in the microplate:

Reagent*	Sample**	Blank
Sample	20 μ l	--
Distilled water	--	20 μ l
Substrate I	170 μ l	170 μ l
Substrate II	10 μ l	10 μ l
Mix, measured at 550 nm and record the absorbance of 10th second and 130th second.		

*Reagents must be added sequentially and should not be premixed prior to addition.

**For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more samples into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.

**For colored samples, we recommend setting a parallel sample background control well with same volume of sample only. Other reagents were replaced by distilled water to the same total volume. Subtract the OD value of the sample background control from the OD value of the sample to correct for interference from the sample's own color.

VI. CALCULATION

Unit Definition: One unit of GalLDH is the amount of enzyme that will reduce 1 μ molCytC per minute.

1.1 According to the protein concentration of sample

$$\text{Activity} = \frac{(\text{OD}_{\text{Sample}(130)} - \text{OD}_{\text{Sample}(10)}) - (\text{OD}_{\text{Control}(130)} - \text{OD}_{\text{Control}(10)})}{\epsilon \times d \times C_{\text{Protein}} \times T} \times \frac{V_{\text{Total}}}{V_{\text{Sample}}} (\text{U/mg})$$

1.2 According to the weight of sample

$$\text{Activity} = \frac{(\text{OD}_{\text{Sample}(130)} - \text{OD}_{\text{Sample}(10)}) - (\text{OD}_{\text{Control}(130)} - \text{OD}_{\text{Control}(10)})}{\epsilon \times d \times W \times T} \times \frac{V_{\text{Total}} \times V_{\text{Assay}}}{V_{\text{Sample}}} (\text{U/g})$$

1.3 According to the quantity of cells or bacteria

$$\text{Activity} = \frac{(\text{OD}_{\text{Sample}(130)} - \text{OD}_{\text{Sample}(10)}) - (\text{OD}_{\text{Control}(130)} - \text{OD}_{\text{Control}(10)})}{\epsilon \times d \times N \times T} \times \frac{V_{\text{Total}} \times V_{\text{Assay}}}{V_{\text{Sample}}} (\text{U}/10^4)$$

1.4 According to the volume of sample

$$\text{Activity} = \frac{(\text{OD}_{\text{Sample}(130)} - \text{OD}_{\text{Sample}(10)}) - (\text{OD}_{\text{Control}(130)} - \text{OD}_{\text{Control}(10)})}{\epsilon \times d \times T} \times \frac{V_{\text{Total}}}{V_{\text{Sample}}} (\text{U/mg})$$

ϵ : molar extinction coefficient, $17.3 \times 10^3 \text{ L/mol/cm} = 17.3 \text{ ml}/\mu\text{mol/cm}$

d : the optical path of 96-Well microplate, 0.6 cm;

C_{Protein} : the protein concentration, mg/ml

W : the weight of sample, g

N : the quantity of cell or bacteria, $N \times 10^4$

V_{Total} : the total volume of all reagents in assay procedure, ml

V_{Sample} : the volume of sample in assay procedure, ml

V_{Assay} : the volume of Assay Buffer II in sample preparation, ml

T : the reaction time, minutes