



Ammonia/Ammonium Colorimetric Microplate Assay Kit User Manual

Catalog # CAK1181

(Version 1.7D)

Detection and Quantification of Ammonia/Ammonium content in
Serum, Plasma, Urine, Saliva, Cell culture, Tissue extracts, Cell lysate,
Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.

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I. INTRODUCTION

Ammonia (NH_3) or its ion form ammonium (NH_4^+) is an important source of nitrogen for living systems. It is synthesized through amino acid metabolism and is toxic when present at high concentrations. In the liver, ammonia is converted to urea through the urea cycle. Elevated levels of ammonia in the blood (hyperammonemia) have been found in liver dysfunction (cirrhosis), while hypoammonemia has been associated with defects in the urea cycle enzymes (e.g. ornithine transcarbamylase). Ammonia/Ammonium Colorimetric Microplate Assay Kit is designed to directly measure NH_3 and NH_4^+ in a variety of samples. In the assay, ammonia reacts with hypochlorous acid, which is determined at 620nm, is directly proportional to the Ammonia/Ammonium concentration in the sample.

II.KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay BufferI	30 mlx 4	4 °C
Assay BufferII	30 mlx 2	4 °C
Dye Reagent I	Powder x 2	4 °C
Dye Reagent II	Powder x 2	4 °C
Dye Reagent II Diluent	3 ml x 1	4 °C
Standard	Powder x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 620 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer

IV. REAGENT PREPARATION

Dye Reagent I: Add 3.5 ml distilled water into 1 bottle Dye Reagent I to dissolve before use. Store at 4 °C and use within 24 hours.

Dye Reagent II: Briefly centrifuge prior to opening. Add 1.5 ml Dye Reagent II Diluent into 1 vial Dye Reagent II, mix before use. Store at 4 °C and use within 24 hours.

Standard: Add 1 ml distilled water to dissolve, then add 5 µl into 995 µl distilled water, mix, the concentration will be 1000 µmol/L. Store at 4°C for 1 month. Perform 2-fold serial dilutions of the top standard solution using distilled water to make the standard curve. The concentration of standard curve could be 1000/500/250/125/62.5/31.2/15.6 µmol/L.

V. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay Buffer I for 5×10^6 cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 10000g 4°C for 10 minutes, take the supernatant into a new centrifuge tube and then add 0.5 ml Assay Buffer II, mix.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay Buffer I, centrifuged at 10000g 4°C for 10 minutes, take the supernatant into a new centrifuge tube and then add 0.5 ml Assay Buffer II, mix.

3. For liquid samples

For serum, plasma, urine and other contain protein samples, add 0.1 ml sample into 0.6 ml Assay Buffer I, centrifuged at 10000g 4°C for 10 minutes, take the supernatant into a new centrifuge tube and then add 0.3 ml Assay Buffer II, mix.

For other sample does not contain proteins, detect directly.

VI. ASSAY PROCEDURE

Add following reagents in the microplate:

Reagent*	Sample**	Standard	Blank
Sample	100 µl	--	--
Standard	--	100 µl	--
Distilled water	--	--	100 µl
Dye Reagent I	70 µl	70 µl	70 µl
Dye Reagent II	30 µl	30 µl	30 µl
Shake and mix, put it into the oven, 37°C for 15 minutes, then record absorbance measured at 620 nm.			

Note:

*Reagents must be added sequentially and should not be premixed prior to addition.

** The concentrations can vary over a wide range depending on the different samples. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.

**For colored samples, we recommend setting a parallel sample background control well with same volume of sample only. Other reagents were replaced by distilled water to the same total volume. Subtract the OD value of the sample background control from the OD value of the sample to correct for interference from the sample's own color.

VII. CALCULATION

1. Calculate the sample concentration in ASSAY PROCEDURE according to the slope of the standard curve

$$C = \frac{(OD_{\text{Sample}} - OD_{\text{Blank}}) - \text{Intercept}}{\text{Slope}} \times n (\mu\text{mol/L})$$

Calculate the initial concentration according to sample preparation procedure.

2. According to one point of the standard OD and concentration

2.1 According to the volume of sample (contain proteins)

$$C = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Blank}})}{(OD_{\text{Standard}} - OD_{\text{Blank}}) \times V_{\text{Sample}}} \times n (\mu\text{mol/ml})$$

2.2 According to the volume of sample (does not contain proteins)

$$C = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Blank}})}{(OD_{\text{Standard}} - OD_{\text{Blank}}) \times V_{\text{Sample}}} (\mu\text{mol/ml})$$

2.3 According to the cell and bacteria of sample

$$C = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Blank}})}{(OD_{\text{Standard}} - OD_{\text{Blank}}) \times (N \times V_{\text{Sample}} / V_{\text{Assay}})} (\mu\text{mol}/10^4)$$

2.4 According to the weight of sample

$$C = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Blank}})}{(OD_{\text{Standard}} - OD_{\text{Blank}}) \times (W \times V_{\text{Sample}} / V_{\text{Assay}})} (\mu\text{mol/g})$$

Slope: the absorbance slope of standard curve

n: the dilution factor

W: the weight of sample, g

N: the quantity of cell or bacteria, $N \times 10^4$

V_{Assay} : the volume of Assay Buffer I and Assay Buffer II, mL

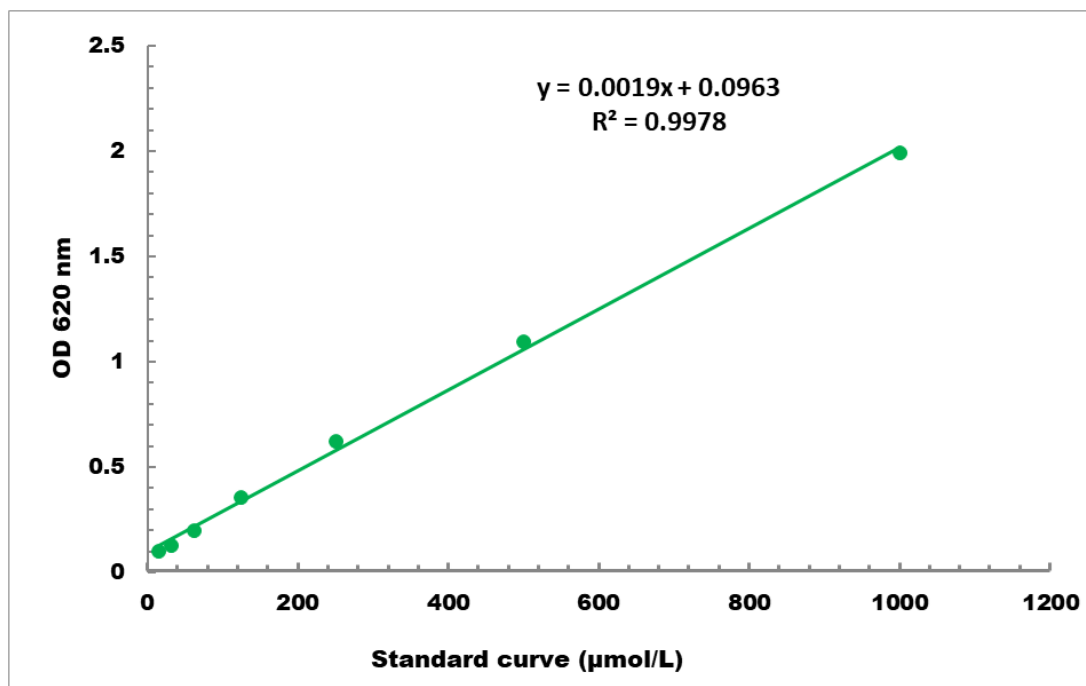
V_{Standard} : the volume of standard in assay procedure, mL

V_{Sample} : the volume of sample in assay procedure, mL

C_{Standard} : the standard concentration, $\mu\text{mol/mL}$

VIII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: $10\mu\text{mol/L}$ - $1000\mu\text{mol/L}$