



Glucose Fluorometric Microplate Assay Kit User Manual

Catalog # CAK8003

(Version 1.2A)

Detection and Quantification of Glucose Content in Serum, Plasma,
Tissue extracts, Cell lysate, Cell culture and Other biological fluids
Samples.

For research use only. Not for diagnostic or therapeutic procedures.

I. INTRODUCTION.....	2
II. KIT COMPONENTS.....	3
III. MATERIALS REQUIRED BUT NOT PROVIDED.....	4
IV. REAGENT PREPARATION.....	5
V. SAMPLE PREPARATION.....	6
VI. ASSAY PROCEDURE.....	7
VII. CALCULATION.....	8
VIII. TYPICAL DATA.....	9

I. INTRODUCTION

Glucose ($C_6H_{12}O_6$) is a key diagnostic parameter for many metabolic disorders.

Increased glucose levels have been associated with diabetes mellitus, hyperactivity of thyroid, pituitary and adrenal glands. Decreased levels are found in insulin secreting tumors, myxedema, hypopituitarism and hypoadrenalism.

Glucose Fluorometric Microplate Assay Kit provides a simple and sensitive method for monitoring glucose content in various samples. In this assay, glucose oxidase specifically oxidizes free glucose generating a compound that reacts with the glucose probe, which can be detected fluorometrically (Ex/Em 535/587).

II.KIT COMPONENTS

Component	Volume	Storage
96-Well Black Microplate	1 plate	
Reaction Buffer	20 mlx 1	4 °C
Enzyme	Powder x 1	-20 °C, keep in dark
Probe	Powder x 1	-20 °C, keep in dark
Probe Diluent	1 mlx 1	4 °C
Standard	Powder x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Fluorescence microplate reader to read fluorescence at Ex/Em = 535/587
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Ice
7. Centrifuge
8. Timer

IV. REAGENT PREPARATION

Standard: Briefly centrifuge prior to opening. Dissolve in 1 ml distilled water to generate 50 mmol/L of standard stock solution, store at 4 °C for 1 month or -20 °C for 1 year after reconstitution. Then dilute to 100 µmol/L standard top solution by adding 2 µl stock solution into 998 µl distilled water. Perform 2-fold serial dilutions of the top standard solution using distilled water to make the standard curve. The concentration of standard curve could be 100/50/25/12.5/6.25/3.12 µmol/L.

Enzyme: Briefly centrifuge prior to opening. Add 1 ml Reaction Buffer to dissolve before use. Aliquot & store at -20 °C. Use within 1 month.

Probe: Warm Probe Diluent to RT prior to use to melt frozen Probe Diluent; Briefly centrifuge prior to opening then add 1 ml Probe Diluent to dissolve. Store at -20 °C, protect from light and moisture. Use within 1 month.

Note: Divide into small aliquots to avoid repeated freeze-thaw cycles.

V. SAMPLE PREPARATION

1. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml distilled water, put it in the boiling water bath for 15 minutes, centrifuged at 8000g 4°C for 10 minutes, take the supernatant into a new centrifuge tube for detection.

2. For liquid samples

Detect directly or dilute with distilled water.

VI. ASSAY PROCEDURE

Warm the solution to room temperature before use.

Add following reagents in the microplate:

Reagent*	Sample**	Standard	Blank
Reaction Buffer	170 μ l	170 μ l	170 μ l
Sample	10 μ l	--	--
Standard	--	10 μ l	--
Distilled water	--	--	10 μ l
Probe	10 μ l	10 μ l	10 μ l
Enzyme	10 μ l	10 μ l	10 μ l
Mix, put it in the oven, 37 °C for 15 minutes, protected from light, record fluorescence measured at Ex/Em = 535/587 nm.			

Note:

*Reagents must be added sequentially and should not be premixed prior to addition.

**The concentrations can vary over a wide range depending on the different samples.

For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.

VII. CALCULATION

1. Calculate the sample concentration in ASSAY PROCEDURE according to the slope of the standard curve

$$C = \frac{(\text{RFU}_{\text{Sample}} - \text{RFU}_{\text{Blank}}) - \text{Intercept}}{\text{Slope}} \times n (\mu\text{mol/L})$$

Calculate the initial concentration according to sample preparation procedure.

2. According to one point of the standard RFU and concentration

2.1 According to the weight of sample

$$C = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{RFU}_{\text{Sample}} - \text{RFU}_{\text{Blank}})}{(\text{RFU}_{\text{Standard}} - \text{RFU}_{\text{Blank}}) \times W \times (V_{\text{Sample}} / V_{\text{Assay}})} (\mu\text{mol/g})$$

2.2 According to the volume of sample

$$C = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{RFU}_{\text{Sample}} - \text{RFU}_{\text{Blank}})}{(\text{RFU}_{\text{Standard}} - \text{RFU}_{\text{Blank}}) \times V_{\text{Sample}}} (\mu\text{mol/ml})$$

Slope: the absorbance slope of standard curve

n: the dilution factor

C_{Standard} : the standard concentration, $\mu\text{mol/L}$

V_{Standard} : the volume of standard in assay procedure, μl

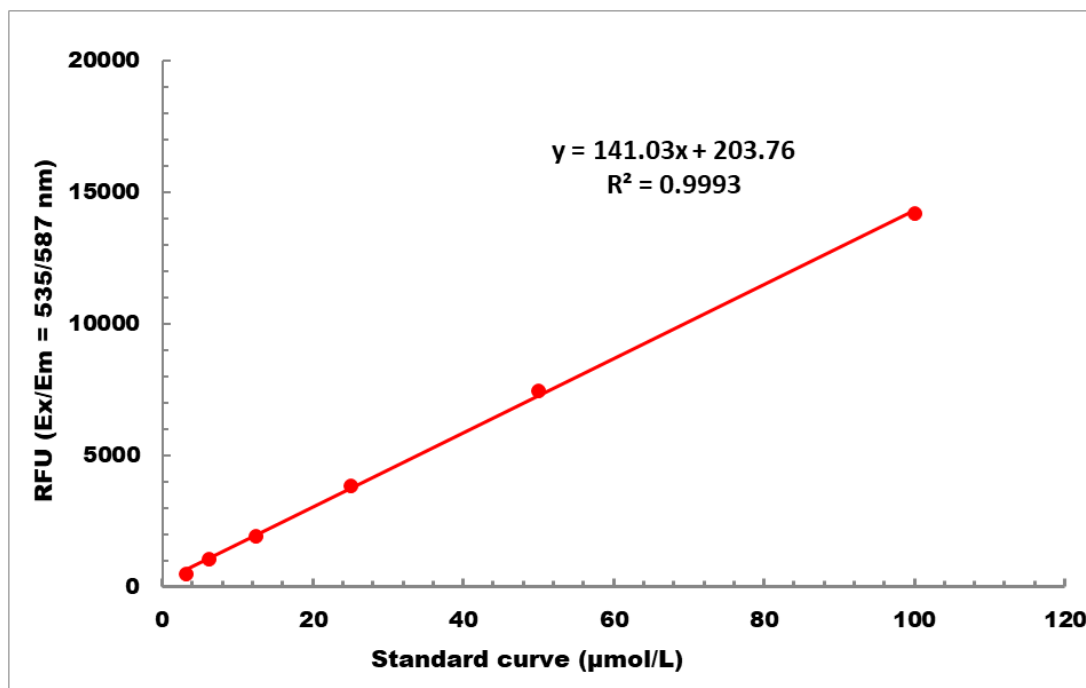
V_{Sample} : the volume of sample in assay procedure, μl

V_{Assay} : the volume of Assay Buffer, μl

W: the weight of sample, g

VIII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 5µmol/L - 100µmol/L